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Bladder cancer stage-associated hub genes revealed by WGCNA co-expression network analysis

Yu Di^{1,2}, Dongshan Chen^{1,2}, Wei Yu³ and Lei Yan^{1*}

Abstract

Background: Bladder cancer was a malignant disease in patients, our research aimed at discovering the possible biomarkers for the diseases.

Results: The gene chip GSE31684, including 93 samples, was downloaded from the GEO datasets and co-expression network was constructed by the data. Molecular complex detection (MCODE) was used to identify hub genes. The most significant cluster including 16 genes: *CDH11*, *COL3A1*, *COL6A3*, *COL5A1*, *AEBP1*, *COL1A2*, *NTM*, *COL11A1*, *THBS2*, *COL8A1*, *COL1A1*, *BGN*, *MMP2*, *PXDN*, *THY1*, and *TGFB111* was identified. After annotated by BiNGO, they were suggested associated with collagen fibril organization and blood vessel development. In addition, the Kaplan Meier curves were obtained by UALCAN. The high expression of *THY1*, *AEBP1*, *CDH11*, *COL1A1*, *COL1A2*, *COL11A1*, *MMP2*, *PXDN*, *BGN*, *COL5A1*, *COL8A1*, and *TGFB111* indicated poor prognosis of the patients ($P < 0.05$). Finally, we examined genes' expression between low and high tumor stage by the Wilcoxon test ($P < 0.05$), *TGFB111* was excluded.

Conclusion: *THY1*, *AEBP1*, *CDH11*, *COL1A1*, *COL1A2*, *COL11A1*, *MMP2*, *PXDN*, *BGN*, *COL5A1*, *COL8A1* associated with the tumor stage as well as tumor patients' prognosis. *COL5A1*, *COL8A1* ($P < 0.01$) may serve as therapeutic targets for the disease.

Keywords: Bladder cancer, Weighted gene co-expression network analysis (WGCNA), Tumor staging, Hub gene identification

Backgrounds

Bladder cancer was ranked fourth of the most common cancers in male, with an estimated 17,240 deaths and 81,190 new cases in 2018 cancer statistics [1]. The survival rate of bladder carcinoma sharply declined with the spreading of the tumor. The most common type of bladder cancer was transitional cell carcinoma, also called urothelial carcinoma. It was often diagnosed at an early stage, producing negative impacts on the patients' daily life and wasting extensive social wealth [2]. Currently, there were four types of standard treatment for bladder cancer including surgery, radiation therapy, chemotherapy, and immunotherapy; surgery was the primary method in clinical trials [3]. Transurethral resection (TUR) with fulguration was suitable for treating superficial bladder

cancer while radical cystectomy was often rendered for patients with bladder cancer at the advanced stage in the situation of which the organs near the bladder such as the prostate and the seminal vesicles of a male patient and the uterus, the ovaries, and part of the vagina of a female need to be removed [4]. Partial cystectomy can be offered to patients who have a low-grade tumor that has invaded the bladder wall but was limited to one area of the bladder [5–7]. Biological therapy was an emerging revolution in the medical field, which involved the use of living organisms, substances derived from living organisms, or laboratory-produced versions of such substances to treat diseases. It can be used to treat cancer itself or the side effects of other cancer treatments [8]. Targeted cancer therapy was a type of biological therapy, which employs targeted-cancer-therapy drugs such as therapeutic antibodies to destroy cancer cells by interfering with a specific molecular target needed for cancer cell growth [9]. As such, it was important to find the critical genes which

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may be related to cancer growth and proliferation and thus can further affect the prognosis of patients. Weighted gene co-expression network analysis (WGCNA) was a systems biology method for describing the correlation patterns among genes across microarray samples, and can be used for finding modules of highly correlated genes, for summarizing such clusters using the module eigengene or an intramodular hub gene, for relating modules to one another and to external sample traits, and then the correlation networks can be used to identify candidate biomarkers or therapeutic targets [10]. It can be used to discover the genes for unknown function with biological processes, or for candidate disease or transcriptional regulatory work. Although it didn't provide causality, the co-expression network can be used to identify regulatory genes underlying various phenotypes [11]. Multiple testing problems inherent in the microarray data analysis were resolved in WGCNA. Network approaches provided means to bridge the gap from individual genes to systems oncology [12]. It was not only suitable for mRNAs, but also for microRNAs and lncRNAs, for example, Giulietti, M et al. [13] have reported the expression of *LINC00675* and *LINC01133* lncRNAs associated with pancreatic cancer's development and progression by co-expression network analysis. Zhou, X. G et al. [14] have found that hsa-miR-125b-5p, hsa-miR-145-5p, hsa-let-7c-5p, hsa-miR-218-5p, and hsa-miR-125b-2-3p were hub miRNAs related to prognosis as well as the pathological stage in colon cancer by the method. In fact, if the data met the requirements, WGCNA can be applied and certain information can be obtained. In our study, the WGCNA algorithm was employed to construct co-expression network, which was integrated with genetic information to explain the biological significance of the module genes, and then identified the hub genes of bladder cancer. Although it has been applied bladder cancer in several articles such as Li, S [15], Deng, S. P. [16], and Gaballah [17], and concluded by Giulietti, M [18], however, they choose to perform WGCNA only on DEG genes which may easily obtain the positive results, reduce the workload of calculation, and at the same time increase the possibility of leaving out the lowly expressed but highly correlated genes. In our research, all the possible genes were analyzed and our results would be more complete and reliable.

Materials and methods

Data collection

The gene chip GSE31684 [19] of bladder cancer with its clinical manifestation data was downloaded from the GEO database (<https://www.ncbi.nlm.nih.gov/geo/>). The platform was GPL570(Affymetrix Human Genome U133 Plus 2.0), and there were 93 bladder cancers samples with several clinical variables. A series of clinical traits are shown in Table 1. The raw data had been processed,

Table 1 The summary clinical information of the samples

Items	Counts value
Gender	
Male	68
Female	25
Age at diagnosis	69.13± 10.15(years)
Survival Months	47.47± 44.52(months)
Stage	
<pt<sub>a</pt<sub>	5
<pt<sub>1</pt<sub>	10
<pt<sub>2</pt<sub>	17
<pt<sub>3</pt<sub>	42
<pt<sub>4</pt<sub>	19
PLND Result	
Negative	49
Positive	28
None	16
Grade	
High	87
Low	6
Distant Lymphonodus Metastasis	
none	59
yes	34
Local Recurrence After Surgery	
0	73
1	20
Metastasis	
0	57
1	36
Smoking	
Former	56
Never	18
Current	19

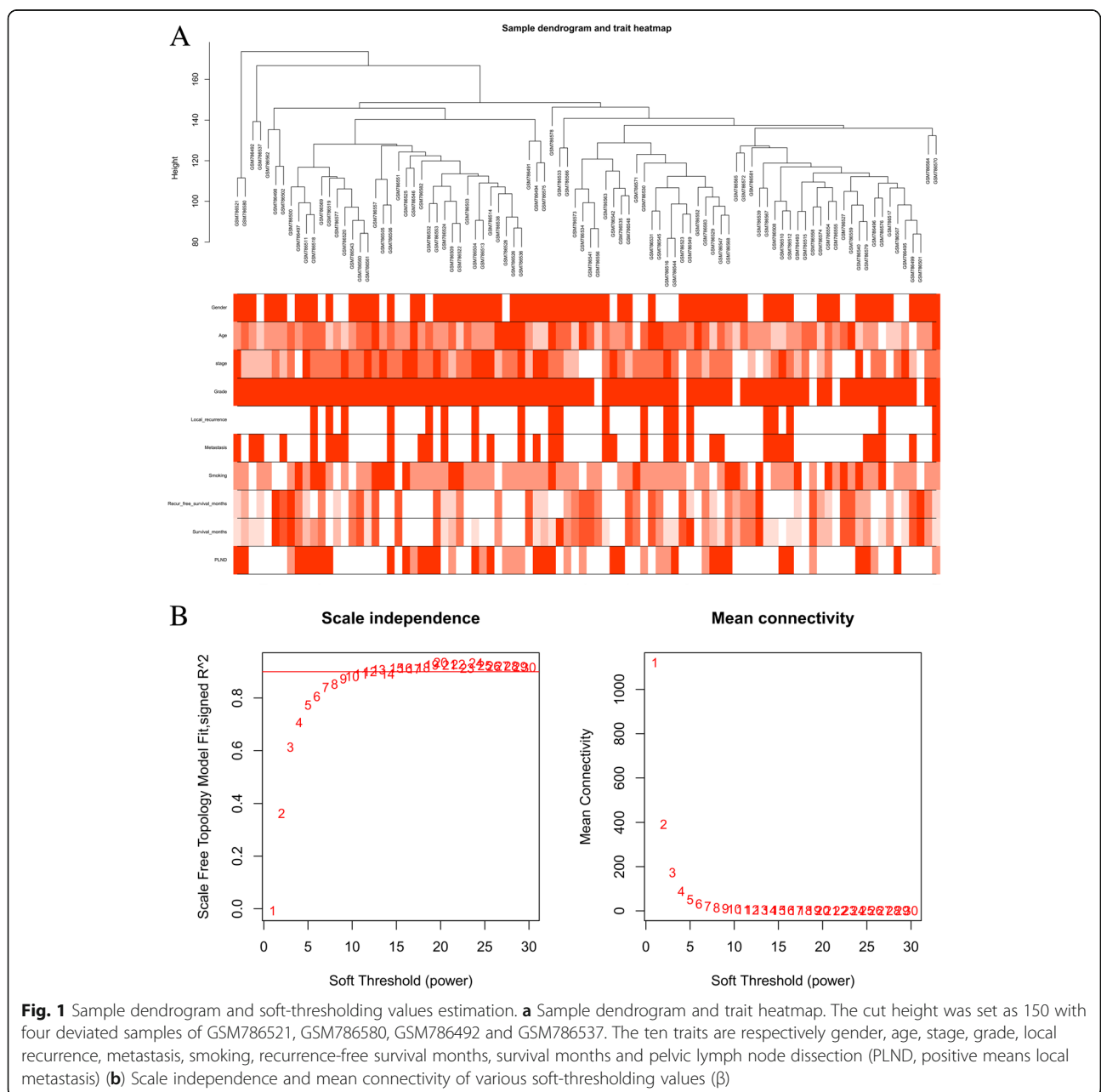
and the gene expression matrix provided by the website was directly used for the analysis. More than 50,000 genes were entered into the BRB-arraytools (<https://lin.us.nci.nih.gov/BRB-ArrayTools/>) [20] for filtering: genes were excluded when less than 20% of expression data had at least a 1.6 -fold change in either direction from gene's median value, and when the percent of data missing or filtered out exceeded 50%. The fold change value is not obtained by comparing the normal and tumor samples. It is a minimum fold-change filter, a parameter in BrB-arraytools, whose detail description can be found in the user manual in the

website (<https://linus.nci.nih.gov/BRB-ArrayTools/Documentation.html>). Finally, a total of 13,222 genes passed the criteria. Compared to analyze the DEGs, our work requires a high equipment computer and carefully distinguish the false positive results, as mentioned above, its merit is obvious: the results would be more complete.

WGCNA analysis of the filtered genes

The “WGCNA” R package was used to construct a co-expression network for the filtered genes [21]. After employing the “hclust” function to the expression matrix evaluated by the average method, gene chips including

GSM786521, GSM786580, GSM786492, and GSM786537 whose cluster height surpass 150 were identified deviated and thus excluded from further analysis (As shown in Fig. 1a). The other 89 samples were used to calculate the Pearson’s Correlation Matrices. The matrix of weighted adjacency was created by formula $a_{mn} = |c_{mn}|^\beta$ (a_{mn} : adjacency between gene m and gene n, c_{mn} : Pearson’s correlation, β :soft-power threshold). Afterward, the clinical trait data were loaded and the scale independence and mean connectivity were estimated. Additionally, the topological overlap measure(TOM) matrix, transformed by the adjacency matrix, was used to estimate its connectivity



property in the network [22]. A hierarchical clustering dendrogram of the TOM matrix was constructed by the average distance with a minimum size threshold of 30 to classify the similar genes expression profiles into different gene modules. The different module eigengenes (MEs) and the clinical traits were then correlated. The gene significance (GS) quantifying associations of individual genes with the clinically interesting trait and the module membership (MM) which acted as the correlation between the module eigengenes and the gene expression profiles were calculated. As proved by previous research, if the GS and MM were highly correlated, the most important (central) elements in the modules were also tightly associated with the trait [23]. As such, they can be used to construct the network and identify the hub genes.

Co-expression network construction and hub genes identification

The Cytoscape v3.6.0 was used to visualize the co-expression network in the module of interest [24]. At the

same time, the network was analyzed by Molecular Complex Detection(MCODE) [25], a Cytoscape plugin that detected densely connected regions in networks that may represent molecular complexes. Algorithms for finding clusters, or locally dense regions, of a graph, were an ongoing research topic in computer science and were often based on network flow/minimum cut theory. The internal mechanism demonstrated that MCODE was suitable for co-expression network, where the proteins in the same complex often displayed high correlations and may be detected by the plugin. The parameters were set as follows. Degree cutoff: 2, node score cutoff: 0.2, cut style: haircut, k-core: 2, and max. Depth: 100. At the same time, WebGestalt was applied to find the transcriptional factors (TFs).

Function annotation of the module of interest and the hub genes

The gene ontology (GO) analysis results of the genes in the module of interest were also provided by

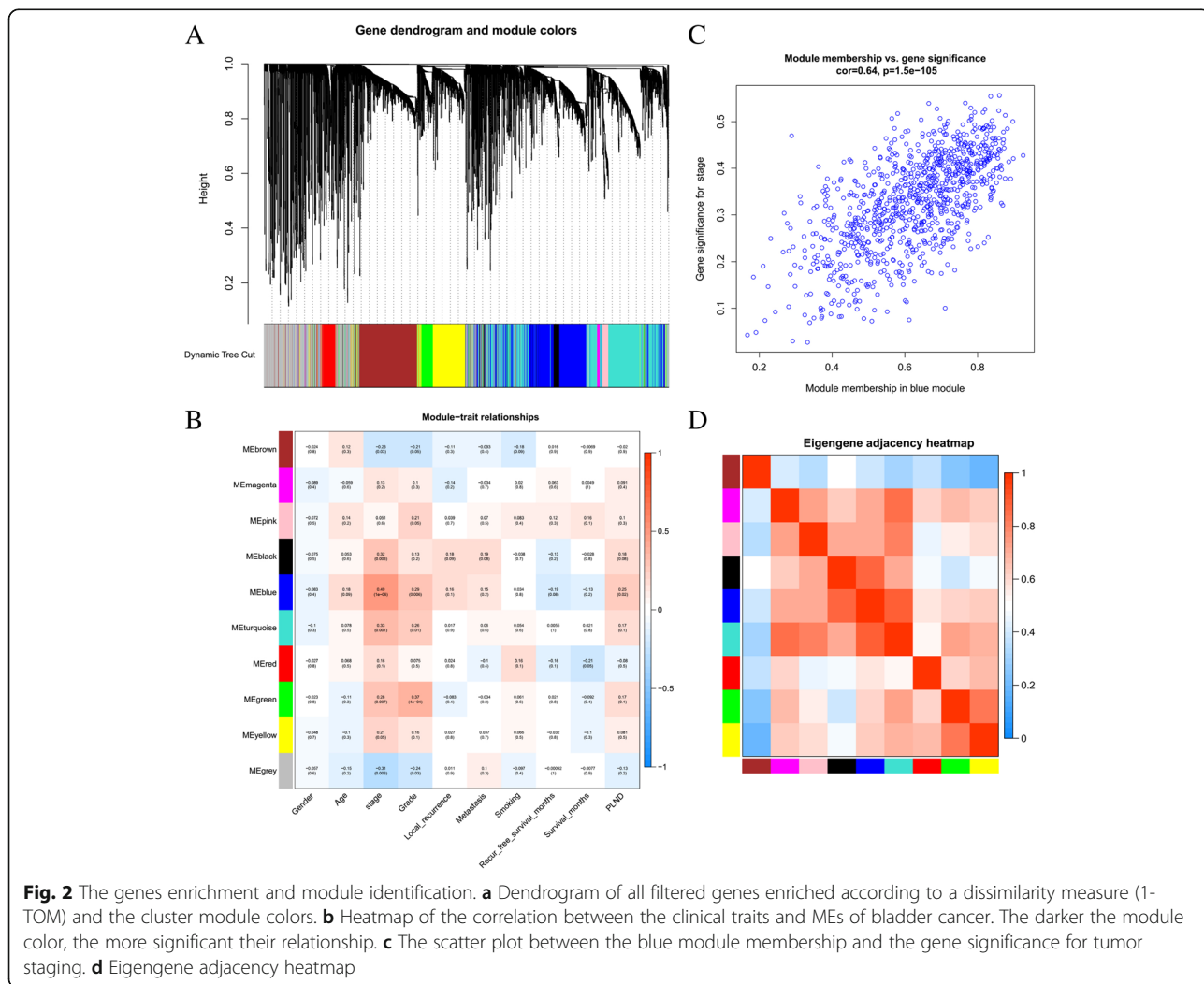


Fig. 2 The genes enrichment and module identification. **a** Dendrogram of all filtered genes enriched according to a dissimilarity measure (1-TOM) and the cluster module colors. **b** Heatmap of the correlation between the clinical traits and MEs of bladder cancer. The darker the module color, the more significant their relationship. **c** The scatter plot between the blue module membership and the gene significance for tumor staging. **d** Eigengene adjacency heatmap

WebGestalt [26]. As a biological network gene ontology tool for functional enrichment analysis in various biological contexts [27], Bingo was used to analyze the hub genes with the following criteria: significant level:0.05; statistic test: binomial; multiple testing corrections: Benjamini and Hochberg False Discovery Rate correction.

Survival analysis by TCGA data and results validation

UALCAN was a web portal for analyzing cancer transcriptome data [28]. It provided easy access to publicly available cancer transcriptome data and allowed users to identify biomarkers or to perform in silico validation of potential genes of interest. The overall survival of bladder cancer was conducted by UALCAN, and the expression levels of the hub gene in different stages were examined.

Results

WGCNA analysis and modules significance calculation

The sample dendrogram and trait heatmap are shown in Fig. 1a. As mentioned above, the samples of GSM786521, GSM786580, GSM786492, and GSM786537 were excluded.

The R package of “WGCNA” was used to classify 13,222 genes with similar expression levels into different modules. In our research, $\beta = 8$ was set to guarantee high scale independence (near 0.9) and low mean connectivity (near 0). The dissimilarity of the modules was set as 0.2, and a total of 10 modules were generated (Fig. 2a). The module trait relationship is shown in Fig. 2b. The blue module associated with the tumor staging was the deepest ($cor = 0.49$, $P = 1E-06$), which was chosen for further analysis (Fig. 2c). The module membership in the blue module and the gene significance have a high correlation (0.64) and high P -value ($1.5E-105$), suggesting that the module is suitable for identifying the hub genes associated with the staging of cancer. The eigengene adjacency heatmap is shown in Fig. 2d, which indicates that the blue module, the black module, and some other modules were adjacent.

Co-expression network construction and hub genes identification

The edges signifying the correlations in the blue module were filtered by a condition of the weight value being

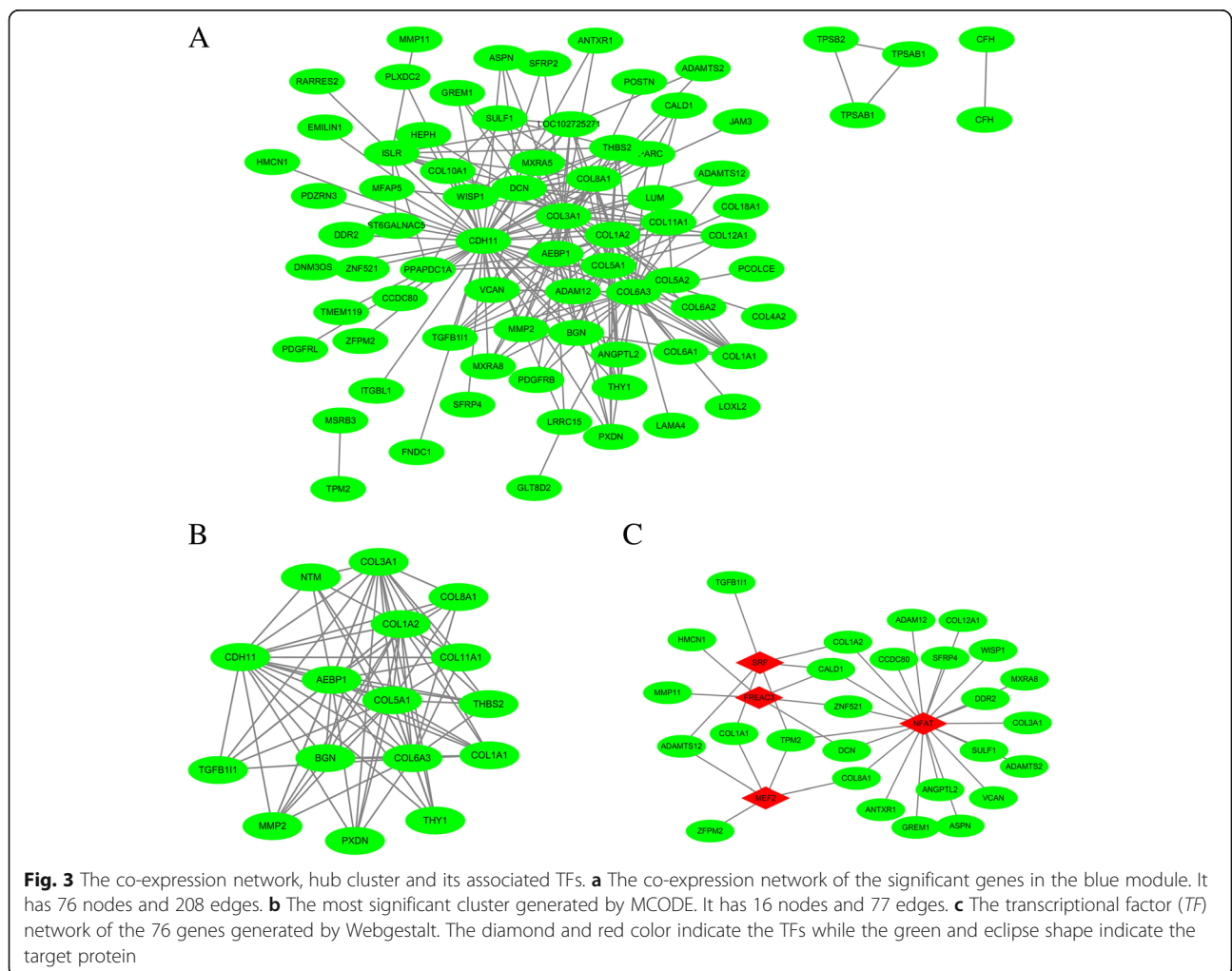
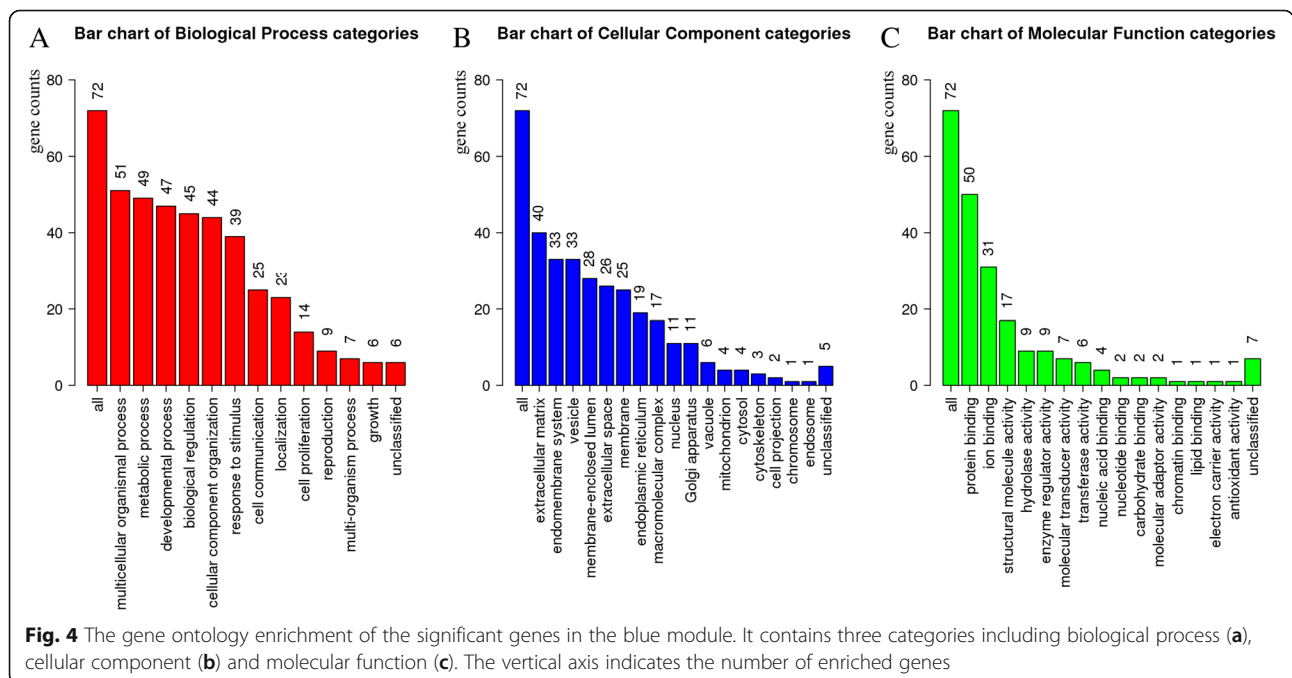


Table 2 The 16 hub genes identified by MCODE (ranked by degree)

Gene Name	Degree	Neighborhood Connectivity	Average Shortest Path Length	Closeness Centrality	Clustering Coefficient	Stress	Topological Coefficient
<i>CDH11</i>	49	6.244898	1.27941176	0.7816092	0.089286	4554	0.115646
<i>COL3A1</i>	35	8.971429	1.5	0.66666667	0.191597	2022	0.133902
<i>COL6A3</i>	31	9.16129	1.55882353	0.64150943	0.197849	1944	0.141633
<i>COL5A1</i>	25	10.72	1.66176471	0.60176991	0.266667	1420	0.16625
<i>AEBP1</i>	21	12.61905	1.70588235	0.5862069	0.347619	852	0.190476
<i>COL1A2</i>	19	14	1.75	0.57142857	0.432749	464	0.212121
<i>NTM</i>	11	19.81818	1.85294118	0.53968254	0.418182	544	0.295794
<i>COL11A1</i>	10	20.7	1.86764706	0.53543307	0.511111	388	0.312121
<i>THBS2</i>	10	21.6	1.86764706	0.53543307	0.488889	432	0.322388
<i>COL8A1</i>	10	21.6	1.86764706	0.53543307	0.488889	432	0.322388
<i>COL1A1</i>	9	21.55556	1.91176471	0.52307692	0.694444	194	0.331624
<i>BGN</i>	8	24.5	1.92647059	0.51908397	0.964286	2	0.376923
<i>MMP2</i>	7	26.85714	1.94117647	0.51515152	1	0	0.413187
<i>PXDN</i>	6	30	1.95588235	0.5112782	1	0	0.461538
<i>THY1</i>	6	30	1.95588235	0.5112782	1	0	0.461538
<i>TGFB111</i>	6	30	1.95588235	0.5112782	1	0	0.461538

greater than 0.2, and a total of 208 edges were obtained. Seventy-six nodes were identified after inputting them into Cytoscape (Fig. 3a). MCODE was applied to discover the hub clusters in the network. The most significant cluster was shown in Fig. 3b. Its degree, neighborhood connectivity, and other parameters

were shown in Table 2. The identified hub genes were *CDH11*, *COL3A1*, *COL6A3*, *COL5A1*, *AEBP1*, *COL1A2*, *NTM*, *COL11A1*, *THBS2*, *COL8A1*, *COL1A1*, *BGN*, *MMP2*, *PXDN*, *THY1*, and *TGFB111*. In addition, we input the 76 genes into the WebGestalt with the P -value < 0.01 and discovered four transcriptional factors including



NFAT, *SRE*, *FREAC3*, and *MEF2*. Its network with the proteins was presented in Fig. 3c.

Function annotation of the module of interest and the hub genes

The results of the GO analysis of the 76 genes in Web-Gestalt were shown in Fig. 4. They were classified into three groups including the biological process, the molecular function, and the cellular component. It was found that the genes were associated with multicellular organism process, extracellular matrix, protein binding and so forth. Table 3 demonstrated the results regarding the hub genes enriched by BiNGO. These genes were suggested related to collagen fibril organization, blood vessel development, cell adhesion, extracellular matrix organization, and collagen biosynthetic process.

Survival analysis and significant gene identification

The bladder cancer expression data in TCGA was analyzed by the UALCAN (<http://ualcan.path.uab.edu/>), and the Kaplan-Meier curves for the overall survival of TCGA patients with bladder cancer were obtained according to the low and high expression of the 16 hub genes. The genes with *P*-values less than 0.01 were *COL5A1*, *COL8A1*, and *TGFB111*(Fig. 5) while those

with *P*-values between 0.05 and 0.01 were *AEBP1*, *CDH11*, *COL1A1*, *COL1A2*, *COL11A1*, *MMP2*, *PXDN*, *BGN*, and *THY1*(Fig. 5). 75% of the genes in the cluster suitable for prognosis prediction of the patients with bladder cancer ($P < 0.05$), suggested that the cluster was important for bladder cancer. The high expression of *COL5A1*, *COL8A1*, *BGN*, *TGFB111*, *AEBP1*, *CDH11*, *COL1A1*, *COL1A2*, *COL11A1*, *MMP2*, *PXDN*, and *THY1* was related to the poor prognosis of the disease. As such, *COL5A1*, *COL8A1* and *TGFB111* can serve as the prognosis biomarkers for patients of bladder cancer. The boxplots demonstrating the correlations between the tumor staging and the hub genes were shown in Fig. 6. The significance of the gene expression in low and high stages was examined by the Wilcoxon test. Except for *TGFB111*, others were significant in distinguishing low and high stage of bladder cancers($P < 0.05$).

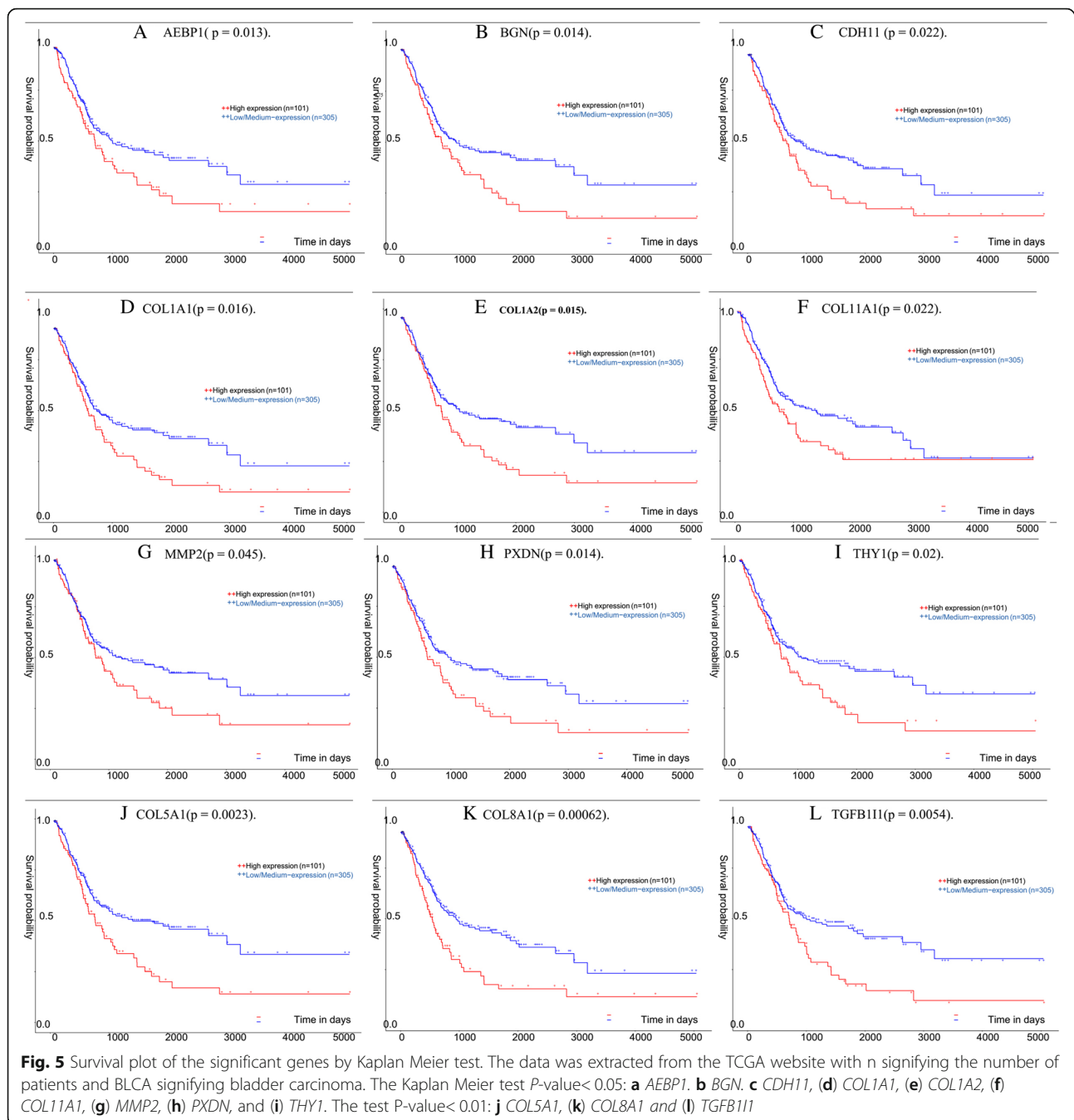
Discussion

Bladder cancer was one of the most common cancers worldwide [1]. Lower-stage tumors were smaller and had a better chance of successful treatment whereas patients with higher-stage cancers may suffer a poor prognosis [29]. With the advancement of science and biological technology, the accuracy treatment of bladder

Table 3 The top 15 GO items of the 16 hub genes provided by BINGO

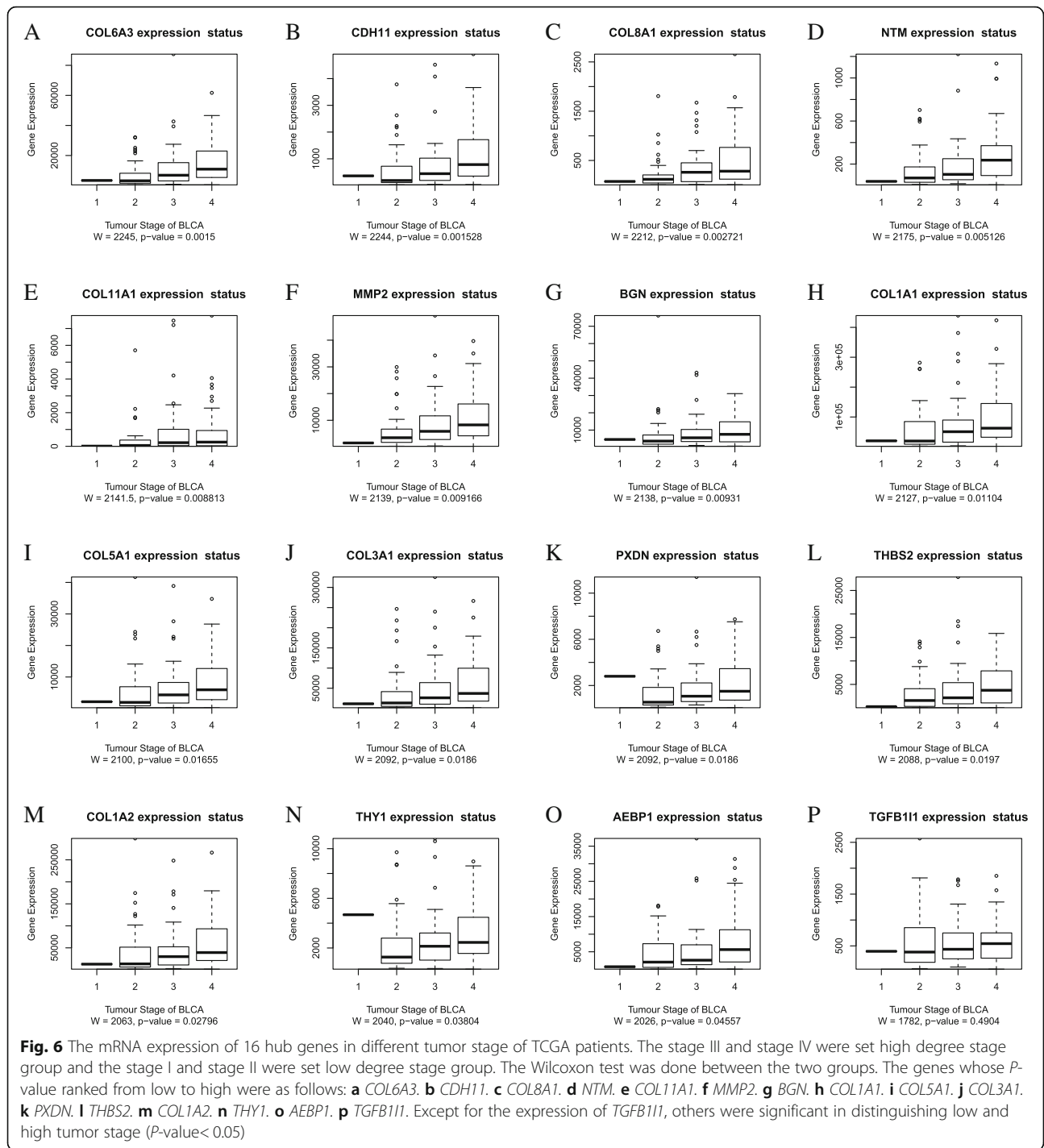
GO-ID	P-value	Description	Genes in the test set
30,199	7.08E-11	collagen fibril organization	<i>COL1A1</i> , <i>COL3A1</i> , <i>COL1A2</i> , <i>COL5A1</i> , <i>COL11A1</i>
1568	7.94E-11	blood vessel development	<i>COL1A1</i> , <i>COL3A1</i> , <i>COL1A2</i> , <i>COL5A1</i> , <i>MMP2</i> , <i>BGN</i> , <i>COL8A1</i> , <i>THY1</i>
7155	2.19E-10	cell adhesion	<i>COL3A1</i> , <i>COL5A1</i> , <i>COL11A1</i> , <i>TGFB111</i> , <i>CDH11</i> , <i>COL6A3</i> , <i>COL8A1</i> , <i>AEBP1</i> , <i>THY1</i> , <i>THBS2</i>
30,198	6.59E-10	extracellular matrix organization	<i>COL1A1</i> , <i>COL3A1</i> , <i>COL1A2</i> , <i>COL5A1</i> , <i>COL11A1</i> , <i>PXDN</i>
32,964	1.94E-08	collagen biosynthetic process	<i>COL1A1</i> , <i>COL3A1</i> , <i>COL5A1</i>
32,963	2.59E-08	collagen metabolic process	<i>COL1A1</i> , <i>COL3A1</i> , <i>COL5A1</i> , <i>MMP2</i>
43,588	4.26E-08	skin development	<i>COL1A1</i> , <i>COL3A1</i> , <i>COL1A2</i> , <i>COL5A1</i>
44,236	7.36E-08	multicellular organismal metabolic process	<i>COL1A1</i> , <i>COL3A1</i> , <i>COL5A1</i> , <i>MMP2</i>
48,513	9.99E-08	organ development	<i>COL1A1</i> , <i>COL3A1</i> , <i>COL1A2</i> , <i>COL5A1</i> , <i>MMP2</i> , <i>TGFB111</i> , <i>BGN</i> , <i>COL6A3</i> , <i>COL8A1</i> , <i>AEBP1</i> , <i>THY1</i>
48,731	1.52E-07	system development	<i>COL1A1</i> , <i>COL3A1</i> , <i>COL1A2</i> , <i>COL5A1</i> , <i>MMP2</i> , <i>TGFB111</i> , <i>CDH11</i> , <i>BGN</i> , <i>COL6A3</i> , <i>COL8A1</i> , <i>AEBP1</i> , <i>THY1</i>
1501	6.53E-07	skeletal system development	<i>COL1A1</i> , <i>COL3A1</i> , <i>COL1A2</i> , <i>MMP2</i> , <i>CDH11</i> , <i>AEBP1</i>
60,346	2.05E-06	bone trabecula formation	<i>COL1A1</i> , <i>MMP2</i>
9653	1.03E-05	anatomical structure morphogenesis	<i>COL1A1</i> , <i>COL1A2</i> , <i>COL5A1</i> , <i>MMP2</i> , <i>TGFB111</i> , <i>BGN</i> , <i>COL8A1</i> , <i>THY1</i>
32,501	1.11E-05	multicellular organismal process	<i>COL11A1</i> , <i>MMP2</i> , <i>TGFB111</i> , <i>BGN</i> , <i>AEBP1</i> , <i>THY1</i> , <i>COL1A1</i> , <i>COL3A1</i> , <i>COL1A2</i> , <i>COL5A1</i> , <i>CDH11</i> , <i>COL6A3</i> , <i>COL8A1</i>
43,589	1.28E-05	skin morphogenesis	<i>COL1A1</i> , <i>COL1A2</i>

Ranked by *P*-value



cancer was gaining momentum, and its development called for the identification of the hub genes relating to bladder cancer. WGCNA was often used to relate modules to external clinical traits and identified the important genes of a tumor. Deng, S. P. et al. [16] had identified “*GDF9*”, “*CYP1A2*”, “*ATF7*”, “*TRPM3*”, “*CER1*”, “*PTPRJ*”, “*KCNIP1*”, and “*LRRC15*” as hub genes in bladder cancer by construction and estimation of two DCNs (normal and cancer state) in 2015. Gaballah [17] identified candidate genes: *PURA*, *SRPK2*, *TRAK1*,

BRD2, and *UPF3* in progression and invasiveness of bladder carcinoma based on DEGs acquired by comparing invasive and noninvasive samples by Limma R packages in 2016. Zhang, X. et al. [30] revealed *POU2F3*, *NKD1*, *CYP2C8*, *LINC00189*, *GCC2* and *OR9Q1* were several remarkable “hub genes” in squamous cell carcinoma of urinary bladder in 2016. The samples were single cells from their own hospital, the DEGs were acquired by NOISeq R packages, and the WGCNA of bladder cancer were focused on intra-tumor heterogeneity



measured by coefficient of variation (CV). Yuan, Lushun et al. [31] have reported that the overexpression of COL3A1 confers a poor prognosis in human bladder cancer in 2017. The DEGs used for WGCNA were obtained by comparing low stage bladder cancers (Ta-T1) and high stage bladder cancers (T2-T4) by Limma R packages, and hub genes were defined by module

connectivity. Li, S. et al. [15] reported *MMP11*, *COL5A2*, *CDC25B*, *TOP2A*, *CENPE*, *CDC43*, *TK1*, *TPX2*, *CDCA8*, *AEBP1*, and *FOXM1* correlated with clinical prognosis of patients with bladder cancer in 2017. The gene chip GSE13507 they used contained 23 recurrent non-muscle invasive tumor tissues, 58 normal looking bladder mucosae surrounding cancer and 10 normal bladder.

Probesets were filtered by their variance across all samples, and hub genes were identified using a network-Screening function. Giulietti, M et al. [18] have reviewed the literature of WGCNA analysis in bladder cancer and concluded hub miRNAs of miR-1-1/2, miR-28, miR-133a-1/2, miR-139, miR-143, miR-195, and miR-6507 in the disease by WGCNA in 2018. However, they choose to perform WGCNA only on DEG genes which may easily obtain the positive results and increase the possibility of leaving out the lowly expressed but highly correlated genes. In the current study, the gene chip GSE31674 including its clinical traits was downloaded from the GEO datasets. In order to avoid the disadvantages and make our results more accurate, all the filtered 13,222 genes were used for WGCNA analysis. We finally obtained 16 hub genes associated the staging of bladder cancer, including *CDH11*, *COL3A1*, *COL6A3*, *COL5A1*, *AEBP1*, *COL1A2*, *NTM*, *COL11A1*, *THBS2*, *COL8A1*, *COL1A1*, *BGN*, *MMP2*, *PXDN*, *THY1*, and *TGFB111*. These genes were associated with the process of collagen fibril organization(GO:0030199)(*COL1A1*,*COL3A1*,*COL1A2*,*COL5A1*,*COL11A1*),blood vessel development(GO:0001568)(*COL1A1*,*COL3A1*,*COL1A2*,*COL5A1*,*MMP2*,*BGN*,*COL8A1*,*THY1*),cell adhesion(GO:0007155)(*COL3A1*,*COL5A1*,*COL11A1*,*TGFB111*,*CDH11*,*COL6A3*,*COL8A1*,*AEBP1*,*THY1*,*THBS2*), extracellular matrix organization(GO:0030198)(*COL1A1*,*COL3A1*,*COL1A2*,*COL5A1*,*COL11A1*,*PXDN*), collagen biosynthetic process(GO:0032964)(*COL1A1*,*COL3A1*,*COL5A1*) in bladder cancer. They were also associated with the pathways of ECM-receptor interaction(*hsa04512*)(*COL3A1*, *COL6A3*, *COL1A2*, *COL1A1*, *COL11A1*, *THBS2*, *COL5A1*), protein digestion and absorption(*hsa04974*)(*COL3A1*, *COL6A3*, *COL1A2*, *COL1A1*, *COL11A1*, *COL5A1*),focal adhesion(*hsa04510*)(*COL3A1*, *COL6A3*, *COL1A2*, *COL1A1*, *COL11A1*, *THBS2*, *COL5A1*),and PI3K-Akt signaling pathway(*hsa04151*)(*COL3A1*, *COL6A3*, *COL1A2*, *COL1A1*, *COL11A1*, *THBS2*, *COL5A1*) in bladder cancer. The expression levels of most of these genes had positive correlations with tumor staging. After examination by the TCGA datasets, *THY1*, *AEBP1*, *CDH11*, *COL1A1*, *COL1A2*, *COL11A1*, *MMP2*, *PXDN*, *BGN*, *COL5A1*, *COL8A1*, and *TGFB111* were associated with the patient's prognosis($P < 0.05$). The P -values of *COL5A1*, *COL8A1*, and *TGF B111* were less than 0.01 and were then thought important among them. However, *TGFB111* was meaningless in revealing tumor stage and was excluded. *COL5A1* [32–35] encoded an alpha chain which was closely related to type XI collagen and was reported contributing to the metastasis of lung adenocarcinoma. *COL8A1* [36–38] was a short α -chains of type VIII collagen, which was connected to angiogenesis and vascular remodeling; it was reported playing an important role in hepatocarcinoma

cells. In our study, we find it was also associated with bladder cancer.

Conclusion

After the weighted gene co-expression network analysis of bladder cancer and the examination of the hub genes, we find *THY1*, *AEBP1*, *CDH11*, *COL1A1*, *COL1A2*, *COL11A1*, *MMP2*, *PXDN*, *BGN*, *COL5A1*, *COL8A1* associated with the tumor stage as well as tumor patients' prognosis. *COL5A1*, *COL8A1* were most significant in the prediction of the bladder cancer's prognosis(P -value < 0.01), and may serve as the therapeutic target for the disease.

Abbreviations

BGN: matrix-remodelling associated 8; BLCA: bladder carcinoma; BP: biological process; CC: cell component; COL5A1: collagen type V alpha 1 chain; COL8A1: collagen type VIII alpha 1 chain; DCNs: differential co-expression networks; GS: gene significance; MF: molecular function; MM: module membership; TGFB111: transforming growth factor beta 1 induced transcript 1; TOM: topological overlap measure; WGCNA: weighted correlation network analysis

Acknowledgments

Thanks for all the authors who provided the technical help for the analysis.

Clinical significance

Our article reports several genes of bladder cancer by Co-expression network analysis, they are associated with tumor's development and patient's prognosis and may serve as the therapeutic targets for the disease, especially in bladder's irrigation.

Funding

This study was supported by the financial grants from the Natural Science Foundation of Shandong Province (Grant nos.2014QLKY16, GG201703180001, and ZR201709230247) (<http://jihx.sdsc.gov.cn/STDPM5/ZR/Default.aspx>).

Availability of data and materials

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

DY contributed to figure construction and article writing while CD and YW helped with drafting the paper, and YL provided fund support and help with the technical problems. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Consent for publication

All the authors have consented for the publication.

Competing interests

The authors declare that they have no competing interest.

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Received: 23 November 2018 Accepted: 17 January 2019

Published online: 28 January 2019

References

- Bray F, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*. 2018;68(6):394–403.
- Habuchi T, et al. Prognostic markers for bladder cancer: international consensus panel on bladder tumor markers. *Urology*. 2005;66(6 Suppl 1):64–74.
- Grayson M. Bladder cancer. *Nature*. 2017;551(7679):S33.
- Chang TC, et al. Image-Guided Transurethral Resection of Bladder Tumors – Current Practice and Future Outlooks. *Bladder Cancer*. 2017;3(3):149–59. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5545914/>. Accessed 27 July 2017.
- Bos MK, et al. Bladder preservation with brachytherapy compared to cystectomy for T1-T3 muscle-invasive bladder cancer: a systematic review. *J Contemp Brachytherapy*. 2014;6(2):191–9.
- Cha, E.K., T.F. Donahue, and B.H. Bochner, Radical transurethral resection alone, robotic or partial cystectomy, or extended lymphadenectomy: can we select patients with muscle invasion for less or more surgery? *Urol Clin North Am*. 2015;42(2):189–viii.
- Gakis G, et al. Systematic review on the fate of the remnant Urothelium after radical cystectomy. *Eur Urol*. 2017;71(4):545–57.
- Smolensky D, Rathore K, Cekanova M. Molecular targets in urothelial cancer: detection, treatment, and animal models of bladder cancer. *Drug Des Devel Ther*. 2016;10:3305–22.
- Nicola AM, et al. Antifungal drugs: new insights in research & development. *Pharmacol Ther*. 2018.
- Langfelder P, Horvath S. WGCNA: an R package for weighted correlation network analysis. *BMC Bioinformatics*. 2008;9:559.
- van Dam S, et al. Gene co-expression analysis for functional classification and gene–disease predictions. *Brief Bioinform*. 2018;19(4):575–92.
- Ghazalpour A, et al. Integrating genetic and network analysis to characterize genes related to mouse weight. *PLoS Genet*. 2006;2(8):e130.
- Giulietti M, et al. LncRNA co-expression network analysis reveals novel biomarkers for pancreatic Cancer. *Carcinogenesis*. 2018.
- Zhou XG, et al. Identifying miRNA and gene modules of colon cancer associated with pathological stage by weighted gene co-expression network analysis. *Onco Targets Ther*. 2018;11:2815–30.
- Li S, et al. Identification of biomarkers correlated with the TNM staging and overall survival of patients with bladder Cancer. *Front Physiol*. 2017;8:947.
- Deng SP, Zhu L, Huang DS. Mining the bladder cancer-associated genes by an integrated strategy for the construction and analysis of differential co-expression networks. *BMC Genomics*. 2015;16(Suppl 3):S4.
- Gaballah H. Integration of Gene Coexpression Network, GO Enrichment Analysis for Identification Gene Expression Signature of Invasive Bladder Carcinoma. Vol. 126; 2016.
- Giulietti M, et al. Emerging biomarkers in bladder Cancer identified by network analysis of transcriptomic data. *Front Oncol*. 2018;8:450.
- Riester M, et al. Combination of a novel gene expression signature with a clinical nomogram improves the prediction of survival in high-risk bladder cancer. *Clin Cancer Res*. 2012;18(5):1323–33.
- Simon R, et al. Analysis of gene expression data using BRB-ArrayTools. *Cancer Inform*. 2007;3:11–7.
- Horvath S, et al. Analysis of oncogenic signaling networks in glioblastoma identifies aspm as a novel molecular target. *USA: Proc Natl Acad Sci*. 2006; 103(46):17402–17407.
- Iancu OD, et al. Cosplicing network analysis of mammalian brain RNA-Seq data utilizing WGCNA and mantel correlations. *Front Genet*. 2015;6:174.
- Zhang B, Horvath S. A general framework for weighted gene co-expression network analysis. *Stat Appl Genet Mol Biol*. 2005;4:Article17.
- Su G, et al. Biological network exploration with Cytoscape 3. *Curr Protoc Bioinforma*. 2014;47:8.13.1–24.
- Bader GD, Hogue CW. An automated method for finding molecular complexes in large protein interaction networks. *BMC Bioinformatics*. 2003;4:2.
- Wang J, et al. WEB-based GENE SeT Analysis toolkit (WebGestalt): update 2013. *Nucleic Acids Res*. 2013;41(Web Server issue):W77–83.
- Maere S, Heymans K, Kuiper M. BiNGO: a Cytoscape plugin to assess overrepresentation of gene ontology categories in biological networks. *Bioinformatics*. 2005;21(16):3448–9.
- Chandrashekar DS, et al. UALCAN: a portal for facilitating tumor subgroup gene expression and survival analyses. *Neoplasia*. 2017;19(8):649–58.
- Dobruch J, et al. Gender and bladder Cancer: a collaborative review of etiology, biology, and outcomes. *Eur Urol*. 2016;69(2):300–10.
- Zhang X, et al. Single-cell analyses of transcriptional heterogeneity in squamous cell carcinoma of urinary bladder. *Oncotarget*. 2016;7(40):66069–76.
- Yuan L, et al. Overexpression of COL3A1 confers a poor prognosis in human bladder cancer identified by co-expression analysis. *Oncotarget*. 2017;8(41): 70508–20.
- Lulinska-Kuklik E, et al. Interactions between COL5A1 gene and Risk of the anterior cruciate ligament rupture. *J Hum Kinet*. 2018;62:65–71.
- Liu W, et al. COL5A1 may contribute the metastasis of lung adenocarcinoma. *Gene*. 2018;665:57–66.
- Pabalan N, et al. Association of COL5A1 gene polymorphisms and risk of tendon-ligament injuries among Caucasians: a meta-analysis. *Sports Med Open*. 2018;4(1):46.
- Lv ZT, et al. Association between polymorphism rs12722 in COL5A1 and musculoskeletal soft tissue injuries: a systematic review and meta-analysis. *Oncotarget*. 2018;9(20):15365–74.
- Zhao Y, et al. siRNA-targeted COL8A1 inhibits proliferation, reduces invasion and enhances sensitivity to D-limonene treatment in hepatocarcinoma cells. *IUBMB Life*. 2009;61(1):74–9.
- Li X, et al. Effects of COL8A1 on the proliferation of muscle-derived satellite. *Cell*. 2018.
- Li S, et al. Co-expression network analysis identified COL8A1 is associated with the progression and prognosis in human Colon adenocarcinoma. *Cell Biol Int*. 2018;63(5):1219–28.

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